ARCIIAR26Br Hammond Alicroplastics, big problems: dissecting effects on the immune system and skeletal repair.
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there is a growing awareness that micro- and nanoplastics, which are increasingly prevalent in our environment, pose risks to human health. This project focuses on the role of microplastics on the innate immune system; particularly neutrophils, which are the body's first line of refense and which are known to engulf microplastics. Our team has data showing that neutrophils change their behaviour when exposed to nicroplastics. We have also identified a novel pro-reparative role for europhils in stabilising fractures. In this project we aim to identify how nicroplastics impact skeletal health in development, injury and regeneration using zebrafish as a model.
While we outline an exemplar project, these is scope for the student to ailor the project to some degree as outlined (in the section on student tersonalisation) below, therefore the objectives could slightly change ollowing the project reading and proposal stage.  In this project we will establish the effects of microplastics on skeletal evelopment, homeostasis and repair and on the cells of the innate mmune system, with a particular focus on neutrophils. In this project we ombine in vitro assays for neutrophil activation, with in vivo work in the ebrafish model and novel coculture techniques established by lammond and Amulic labs in which we can culture human neutrophils in zebrafish scales (which are miniature bone organs). Co-supervisor torko Amulic has established that microplastics induce transcriptional changes in human neutrophils, changing multiple inflammatory genes. imilar effects have been demonstrated in the zebrafish model, in which dult zebrafish treated with microplastics (polyethelene and olystyrenes) show changes to inflammatory cytokines. In vitro assays performed in skeletal cells (osteoblasts) show that microplastics can inhibit osteoblast maturation, can induce cellular stress and therefore mpact ability of the cells to form bone. Our lab have shown that eutrophils play a key role in mediating fracture repair, and we and our ollaborators have identified pathways which could mediate neutrophil to osteoblast interactions.  In this project we will bring these findings together to test how incroplastics affect skeletal repair using the genetically tractable ebrafish model. Zebrafish offer excellent opportunities to dissect ellular responses to environmental pollutants as they are optically ranslucent and have reporter lines for many cell types of interest llowing us to interrogate the whole system.  One of the key objectives are to: Using larvae, establish whether mode of delivery (injection, incubation or ingestion) affect microplastic uptake and effects on a) neutrophils and b) skeletal cells
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To test whether acute delivery of microplastics has same effects as chronic delivery on fracture repair. We will test:

- Recruitment of neutrophils to site of injury (dynamic imaging of transgenic reporters)
- Degree of ROS production and NETosis at site of injury (live imaging of dyes such as CellRox and of transgenic reproters)
- Reverse migration/resolution of immune response
- Osteoblast recruitment (using transgenic reporters)
- Bone formation (live alizarin red and calcein staining)
- Quality of bone repair (microCT, nanoindentation, histology, atomic force microscopy)

To dissect whether the primary effect on skeletal outcomes is driven by neutrophils or skeletal cells by using coculture systems (human neutrophils on fish bone) by:

- Incubating human neutrophils with microplastics then exposing them to skeletal injury on untreated fish scales
- Incubating untreated human neutrophils with injured fish scales treated with microplastics prior to injury.

For both assays we will measure neutrophil migration, activation and skeletal repair.

The project as a whole will give the student a highly desirable cross disciplinary skill set, and should be highly publishable

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